

Algal flora of subalpine peat bog pools in the Krkonoše Mts

Řasy tůně krkonošských subalpínských rašelinišť

Sylvie Nováková

Department of Botany, Charles University, Benátská 2, CZ-128 01 Praha 2, Czech Republic, e-mail: sylnova@natur.cuni.cz

Nováková S. (2002): Algal flora of subalpine peat bog pools the Krkonoše Mts. – Preslia, Praha, 74: 45–56.

During a two-year investigation of the Úpské rašeliniště peat bog and the Pančavské rašeliniště peat bog in the Krkonoše Mts (Czech Republic) about 228 taxa of cyanobacteria and algae were found. The diatoms were the dominant group in most of the investigated samples. The relationship between algal flora and the environmental characteristics of the pools was studied. The pH, shading and type of bed were most important factors influencing algal communities in the pools.

Key words: algae, peat bog, Krkonoše Mts, ecology

Introduction

The first phycological records from the Krkonoše Mts were published by Zacharias and his associates (Zacharias 1896, 1898, Lemmermann 1896, Müller 1898, Schröder 1898) but only Lemmermann studied algae in the peat bogs. Also Beck-Managetta (1927, 1929) and Pochmann (1940) collected algae from various sites in the Krkonoše Mts, including peat bogs. Kalina (1969a, 1969b, 1970) described new taxa, i.e. *Gloeochrysis montana* Kalina, *Mallomonas corcontica* (Kalina) Péterfi, *Exanthemachrysis (Corcontochrysis) noctivaga* (Kalina) Gayral, from the “Úpské rašeliniště” peat bog. Recently a research project devoted to the silica-scaled chrysophytes of the Czech Republic has been carried out (Němcová et al. 2001). In contrast to sporadic research on the Czech side of the border in the last fifty years, Matula (1980a, 1980b, 1995 etc.) was intensively concerned about the taxonomy and ecology of algae in the Polish Karkonosze Mts.

Localities

The studied peat bogs occur at two summit plateaus above the timberline, which are important spring areas (the Elbe, Pančava, Úpa, and White Elbe rivers). The Pančavské rašeliniště peat bog ($50^{\circ}45'55''N$, $15^{\circ}32'30''E$) lies at an average altitude of 1335 m and occupies an area of 26 ha. The Úpské rašeliniště peat bog ($50^{\circ}44'10''N$, $15^{\circ}42'40''E$) extends on both sides of the Czech-Polish border at an average altitude of 1425 m and its area is about 10 ha.

Both these raised bogs differ from mires occurring in other Central European mountains and according to phytosociology and microtopography, they resemble bogs in the Alps and Scandinavian peatlands (Rudolph et al. 1928, Jeník & Soukupová 1992). There are two soil types in the area: loamy soils of a humic podzol type and shallow peat soils

(Mejstřík & Straka 1964). The vegetation consists of oligotrophic communities belonging to the *Oxycocco-Sphagnetea*, and of minerotrophic fen communities of the class *Scheuchzerio-Caricetea fuscae* (Jeník & Soukupová 1992).

Fourteen pools at the Pančavské rašeliniště and seven pools at the Úpské rašeliniště were selected for examination. In trying to record the total range of variation of the peat bog waterbodies, more than just characteristic peaty pools were studied. The deeper pools in the peat bog area, whose beds reach the loamy soil because of a very thin peat layer, and pools in streams were examined too.

Material and methods

Samples of water were taken in 4–6-week intervals from May 1998 to October 1999. Submerged mosses and other plants and surface sediments of the pools were also collected to obtain metaphytic and benthic algae. At the same time the water temperature, pH and conductivity were measured by the conductivity/pH meter Gryf 107.

Most of the taxa were identified directly from fresh samples, although some species must have been cultivated. The diatoms and silica-scaled chrysophytes were examined after preparation according to Kramer & Lange-Bertalot (1986). A light microscope was used to determine the majority of the taxa; the scale-bearing chrysophytes were determined by the transmission electron microscope Philips 300.

The presence of species in samples and the environmental data (Table 1) were analysed by multivariate statistical methods using the programme CANOCO (ter Braak & Šmilauer 1998).

Table 1. – Characteristics of the pools studied. Locality: PR – Pančavské rašeliniště, UR – Úpské rašeliniště; Shading: 0 – no shade, 1 – partly shaded, 2 – intensive shade; Flow: 0 – lentic biotope, 1 – lotic biotope, bed: 0 – peaty, 1 – loamy. Note: ⁺ very changeable, ⁺⁺ impossible to measure maximum depth

Sampl. site	Locality	Area (m ²)	Relative depth (%)	Shadowing	Flow	Bed	pH	Conductivity (µS·cm ⁻¹)
1	PR	3.5	2.37	1	0	0	3.9–4.5	12–41
2	PR	0.8	6.93	2	0	0	4–4.3	27–38
3	PR	0.1	19.61	2	0	0	3.8–4	35–52
4	PR	0.8	6.93	2	0	0	3.8–4.3	31–51
5	PR	2.5	33.62	0	0	0	4.4–5.4	20–32
6	PR	1.5	14.47	1	0	0	5–5.3	20–27
7	PR	1.8	13.21	0	0	0	4.4–5.7	25–29
8	PR	450	> 2.4 ⁺⁺	0	0	0	3.6–5.4	7–31
9	PR	0.5	18.79	0	1	1	3.4–5.5	13–55
10	PR	6.5	13.9	0	0	1	3.5–4.2	30–57
11	PR	2	6.26	0	0	0	3.7–4.6	18–69
12	PR	5	1.19	0	1	0	4.1–5.2	15–24
13	PR	4	3.54	0	0	0	3.7–4.5	21–34
14	PR	3	40.92	0	1	1	4.1–5.6	21–29
15	UR	55	1.79	0	1	1	4.4–6.4	21–45
16	UR	2.5	> 28 ⁺⁺	0	0	1	5.2–6.2	14–37
17	UR	4	6.65	1	0	0	3.9–4.6	24–42
18	UR	17	1.07	0	0	0	3.8–4.6	19–91
19	UR	± 200 ⁺	± 1.3	0	0	0	5.8–6.8	15–116
20	UR	1100	> 1.4 ⁺⁺	0	0	0	3.8–4.8	7–38
21	UR	30	> 4.9 ⁺⁺	0	0	0	3.9–4.6	11–42

Results

Two hundred and twenty eight species of cyanobacteria and algae were found (Table 2). The most frequent group of algae were diatoms (*Bacillariophyceae*, 68 species); desmids (*Conjugatophyceae*, 51 species) were the second most dominant group. *Pinnularia viridis*, *Frustulia rhomboides*, *Pinnularia subcapitata*, members of the genus *Eunotia*, *Chroococcus turgidus* and *Cylindrocystis brebissonii* belonged to the most frequently found species.

Table 2. – Distribution of species in the pools studied. Occurrence in sampling sites (expressed as the number of samples in which the species occurred), total occurrence and percentage of all samples in which the species appeared are presented. The system of classes follows van den Hoek et al. (1995). See Table 1 for characteristics of sampling sites. The species codes correspond to Fig. 3.

Sampling site (number of samples taken)	Code	1 (7)	2 (6)	3 (5)	4 (5)	5 (7)	6 (7)	7 (6)	8 (9)	9 (7)	10 (7)	11 (9)	12 (9)	13 (8)	14 (8)	15 (11)	16 (10)	17 (10)	18 (9)	19 (10)	20 (11)	21 (11)	Total (172)	Total (%)
<i>Cyanophyceae</i> :																								
<i>Anabaena inaequalis</i> (Kütz.) Bornet et Flahault	ANBINQ	–	–	–	–	–	–	–	–	–	–	9	3	–	–	8	–	–	20	11.6				
<i>Aphanocapsa hyalina</i> (Lyngbye) Hansg.	APCHYA	2	1	1	4	–	–	2	4	6	2	6	–	2	5	5	4	5	6	55	32.0			
<i>Aphanothecce microscopica</i> Nüg.		–	–	–	–	–	–	–	–	–	–	–	6	1	–	–	3	–	–	10	5.8			
<i>Aphanothecce</i> sp.		–	–	–	–	–	–	1	–	–	2	4	–	–	–	–	–	–	–	–	7	4.1		
<i>Chroococcus turgidus</i> (Kütz.) Nüg.	CHRTUR	7	–	1	7	6	3	7	3	2	9	9	8	4	2	1	10	9	4	10	11	113	65.7	
<i>Cyanothecce cf. aeruginosa</i> (Nüg.) Komárek		–	–	–	–	–	–	–	–	–	1	–	–	–	–	–	–	–	–	–	–	1	0.6	
<i>Geitlerinema cf. amphibium</i> (Ag. ex Gom.) Anagn.		–	–	–	–	–	–	4	–	–	5	5	2	5	2	–	–	5	3	1	32	18.6		
<i>Geitlerinema splendidum</i> (Grev. ex Gom.) Anagn.		–	–	–	–	–	–	–	–	–	–	–	–	–	–	–	–	–	6	–	6	3.5		
<i>Gloeocapsa</i> sp.		2	–	–	–	–	–	1	–	5	4	4	2	–	1	1	6	3	1	2	32	18.6		
<i>Merismopedia glauca</i> (Ehrenb.) Kütz.		–	–	–	–	–	4	–	–	1	1	–	6	–	10	6	5	4	4	41	23.8			
<i>Phormidium cf. animale</i> (Ag. ex Gom.) Anagn.		–	–	–	–	–	–	–	–	–	–	–	–	–	–	–	3	–	–	3	–	1.7		
<i>Phormidium formosum</i> (Bory ex Gom.) Anagn.		–	–	–	–	–	–	–	–	7	2	–	–	–	–	2	–	–	11	6.4				
<i>Phormidium tergestinum</i> (Kütz. ex Gom.) Anagn.	PHOTER	–	–	–	–	–	–	–	–	–	–	–	–	1	–	–	9	–	–	10	5.8			
<i>Pseudanabaena cf. catenata</i> Lauterborn	PSACAT	–	–	–	–	3	–	–	–	–	–	–	1	–	–	3	–	2	9	5.2				
<i>Snowella</i> sp.	SNOWSP	1	–	–	–	–	–	4	–	–	–	3	–	–	2	–	1	11	6.4					
<i>Rhodophyceae</i> :																								
<i>Batrachospermum</i> sp.		–	–	–	–	–	–	1	–	–	–	–	–	–	–	–	–	–	–	–	1	0.6		
<i>Dinophyceae</i> :																								
<i>Peridinium umbonatum</i> Stein	PERUMB	1	–	–	1	–	–	1	–	4	3	1	–	1	2	1	3	3	3	24	14.0			
cysts		2	–	–	1	–	–	–	1	2	–	1	–	–	1	2	3	4	4	21	12.2			
<i>Cryptophyceae</i> :																								
<i>Chilomonas oblonga</i> Pascher	CHIOBL	3	1	1	2	–	–	2	–	1	2	6	7	–	–	2	7	4	6	44	25.6			
<i>Cryptomonas cylindrica</i> Ehrenb.	CRYCYL	1	2	–	3	–	–	3	–	1	1	2	1	2	1	–	–	2	–	1	–	20	11.6	
<i>Cryptomonas reflexa</i> Skuja		–	–	–	–	–	3	–	2	–	–	–	3	2	1	5	–	1	–	2	19	11.0		
<i>Cryptomonas obovata</i> Skuja	CRYBO	1	1	–	1	–	–	5	6	4	1	1	3	1	–	5	4	3	5	41	23.8			
<i>Cryptomonas phaseolus</i> Skuja		–	–	–	–	–	–	–	–	–	–	–	1	–	–	–	–	–	–	1	0.6			
<i>Cryptomonas pyrenoidifera</i> Geitler		–	–	–	–	1	–	–	–	–	–	–	1	–	1	–	1	–	–	4	2.3			
<i>Cryptomonas</i> sp. 1	CRYSP1	1	–	2	–	1	–	–	3	–	1	–	1	–	–	1	–	1	1	12	7.0			
<i>Cryptomonas</i> spp.		2	3	1	2	4	4	3	1	–	4	3	3	4	5	3	1	3	3	2	7	6	64	37.2
<i>Chrysophyceae</i> :																								
<i>Chrysocapsa</i> sp.		–	–	–	–	2	–	–	–	–	–	–	–	–	–	–	–	1	–	3	1.7			
<i>Chrysococcus triporus</i> Matv.		–	–	–	–	–	–	–	–	–	–	3	2	–	–	6	3	–	14	8.1				
<i>Dinobryon divergens</i> Imhof	CRSTRI	–	–	–	2	–	–	8	3	–	6	–	3	1	–	2	8	–	7	–	40	23.3		
<i>Mallomonas calcicola</i> Bradley	DINDIV	–	–	–	–	1	–	–	–	–	–	1	–	–	–	–	–	–	–	2	1.2			
<i>Mallomonas crassissquama</i> (Asmund) Fott		–	–	–	–	1	–	–	–	–	–	–	–	–	–	–	–	–	1	0.6				
<i>Mallomonas flora</i> Harris et Bradley		–	–	–	–	–	–	–	–	–	–	–	1	–	–	–	–	–	1	0.6				
<i>Mallomonas papillosa</i> Harris et Bradley		–	–	–	–	2	–	–	–	–	–	–	–	–	–	–	–	–	2	1.2				
<i>Mallomonas pillula</i> Harris		–	–	–	–	–	–	–	–	–	–	–	2	–	–	–	–	–	2	1.2				
<i>Mallomonas</i> spp.		–	–	–	3	1	2	–	–	2	4	4	2	3	1	–	3	–	2	–	27	15.7		
<i>Ochromonas</i> sp.		1	–	–	1	–	–	–	1	2	–	1	1	3	3	5	2	–	1	–	21	12.2		
<i>Paraphysomonas vestita</i> (Stokes) De Saedeleer		–	–	–	–	1	–	–	–	–	–	–	–	–	–	–	–	–	–	1	0.6			
<i>Synura echinulata</i> Korshikov		–	–	–	–	2	1	–	–	–	2	–	–	3	–	–	2	–	–	10	5.8			
<i>Synura petersenii</i> Korshikov		–	–	–	–	1	–	–	–	–	–	1	2	–	–	–	–	–	4	2.3				
<i>Synura sphagnicola</i> Korshikov		–	–	–	–	–	–	–	1	–	–	2	–	–	–	–	–	–	3	1.7				
<i>Synura</i> spp.		–	–	–	3	1	2	–	–	4	1	2	–	2	1	–	1	–	–	17	9.9			
<i>Bacillariophyceae</i> :																								
<i>Achnanthes</i> cf. <i>lanceolata</i> (Bréb.) Grunov	ACHLAN	–	–	–	–	–	–	1	1	–	–	–	5	2	–	–	–	1	10	5.8				
<i>Achnanthes minutissima</i> Kütz.		–	–	–	1	4	4	–	3	1	3	2	4	6	9	3	1	–	2	47	27.3			

Sampling site (number of samples taken)	Code	1 (7)	2 (6)	3 (5)	4 (5)	5 (7)	6 (7)	7 (6)	8 (9)	9 (7)	10 (7)	11 (9)	12 (9)	13 (8)	14 (8)	15 (11)	16 (10)	17 (10)	18 (9)	19 (10)	20 (11)	21 (11)	Total (172)	Total (%)		
<i>Achnanthes scotica</i> Flower	ACHSCO	—	—	—	—	—	7	1	—	—	1	5	7	5	—	1	2	—	29	16.9						
<i>Amphora</i> sp.		—	—	—	—	—	—	—	—	—	—	—	—	—	—	1	1	—	2	1.2						
<i>Anomooneis serians</i> (Bréb.) Grunov	ANOBRA	—	—	—	6	7	—	—	1	—	2	9	5	4	—	—	3	—	—	37	21.5					
<i>Anomooneis serians</i> (Bréb.) Cleve	ANOSER	1	—	—	—	—	—	—	1	2	—	8	1	3	2	1	—	1	1	—	1	22	12.8			
<i>Aulacoseira ambigua</i> (Grunov) Simonsen		2	—	—	—	—	—	—	—	—	—	2	1	—	—	—	1	1	1	8	4.7					
<i>Aulacoseira distans</i> (Ehrenb.) Simonsen	AULDIS	—	—	—	3	6	2	—	1	—	1	3	6	10	9	1	3	1	1	48	27.9					
<i>Aulacoseira pfaaffiana</i> (Reinisch) Krammer		—	—	—	—	—	—	—	—	—	—	—	1	—	—	—	—	—	—	1	0.6					
<i>Caloneis bacillina</i> (Grunov) Cleve		—	—	—	—	—	—	1	6	—	—	—	—	—	—	—	—	—	—	7	4.1					
<i>Caloneis silicula</i> (Ehrenb.) Cleve	CALSIL	—	—	—	—	—	—	—	—	—	—	—	—	10	4	—	—	—	—	14	8.1					
<i>Caloneis</i> sp.		—	—	—	—	—	—	—	—	—	—	—	—	1	—	—	—	—	—	—	2	1.2				
<i>Cocconeis placenta</i> Ehrenb.		—	—	—	—	—	—	—	—	—	—	—	—	2	2	—	1	—	—	—	5	2.9				
<i>Cyclotella cf. radiosa</i> (Grunov) Lemmerm.		—	—	—	—	—	—	—	—	—	—	1	—	—	2	1	—	1	1	7	4.1					
<i>Cymatopleura</i> sp.		—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	1	0.6					
<i>Cymbella amphicephala</i> Nág.		—	—	—	—	—	—	—	—	1	1	—	—	11	10	—	2	10	—	1	36	20.9				
<i>Cymbella gracilis</i> (Ehrenb.) Kütz.	CMBAMP	—	—	—	4	4	—	1	—	—	1	2	10	9	—	3	—	—	34	19.8						
<i>Cymbella hebridica</i> (Grunov) Cleve	CMBGRA	—	—	—	—	—	—	—	—	—	—	8	3	1	—	—	—	—	20	11.6						
<i>Cymbella minutula</i> Hilde	CMBHEB	—	—	—	7	1	—	—	—	—	—	—	8	3	1	—	—	—	—	2	43	25.0				
<i>Diatomus mesodon</i> (Ehrenb.) Kütz.	CMBMIN	—	—	—	—	4	—	—	—	1	1	2	—	10	1	2	9	—	—	29	16.9					
<i>Diploneis elliptica</i> (Kütz.) Cleve	DIAMES	1	—	—	—	6	—	—	1	1	—	—	8	9	—	1	—	—	—	12	7.0					
<i>Diploneis puella</i> (Schumann) Cleve	DIPELI	—	—	—	—	—	—	—	—	—	—	—	7	3	—	2	—	—	—	1	0.6					
<i>Epithemia</i> sp.		1	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	4	2.3					
<i>Eunotia arcus</i> Ehrenb.		—	—	—	—	—	—	—	—	—	—	—	—	6	—	—	—	—	—	1	—	7	4.1			
<i>Eunotia bilunaris</i> (Ehrenb.) Mills	EUNBIL	3	—	—	7	7	4	5	4	1	3	9	8	8	11	10	—	5	9	2	6	102	59.3			
<i>Eunotia exigua</i> s.l. (Ehrenb.) Rabenh.	EUNEXI	5	3	4	4	6	6	3	3	2	7	7	8	8	5	2	1	8	8	3	11	9	113	65.7		
<i>Eunotia glacialis</i> Meister		1	—	—	7	7	4	—	2	5	—	9	8	4	4	2	2	1	—	1	57	33.1				
<i>Eunotia lapponica</i> Grunow		2	—	—	2	—	—	—	3	9	5	7	2	1	9	8	2	—	2	52	30.2					
<i>Eunotia meisteri</i> Hust.		—	—	—	—	—	—	—	—	1	—	—	2	7	1	—	—	—	—	11	6.4					
<i>Eunotia nymanniana</i> Grunow		—	—	—	3	—	—	—	—	—	8	8	8	3	2	—	1	3	1	37	21.5					
<i>Eunotia praeputia</i> Ehrenb.		—	—	—	6	—	—	1	3	—	2	4	1	1	—	1	1	—	—	22	12.8					
<i>Eunotia rhomboidea</i> Hust.	EUNRHO	—	—	—	7	7	3	—	1	3	8	5	5	—	6	—	4	1	—	—	50	29.1				
<i>Eunotia soleirolii</i> (Kütz.) Rabenh.	EUNSOL	—	—	—	2	6	6	—	1	2	1	1	2	1	2	11	10	4	1	1	—	5	32.0			
<i>Eunotia tenella</i> (Grunow) Hust.	EUNTEN	6	—	—	7	7	6	1	5	5	7	9	8	8	11	10	7	6	3	3	8	117	68.0			
<i>Eunotia</i> spp.		2	—	—	—	—	—	—	2	2	2	2	2	2	2	—	—	—	—	2	—	14	8.1			
<i>Fragilaria constricta</i> Ehrenb.	FRGVIR	4	—	—	2	7	6	2	1	4	2	1	7	11	10	—	2	1	2	1	64	37.2				
<i>Fragilaria virescens</i> Ralfs		—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	2	1.2					
<i>Fragilaria</i> sp.		—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	6	3.5					
<i>Frustulia rhomboides</i> (Ehrenb.) De Toni	GOMPAR	1	—	—	—	—	—	—	—	6	—	2	2	2	2	—	—	—	—	—	30	17.7				
<i>Gomphonema gracile</i> Ehrenb.		—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	1	0.6				
<i>Gomphonema parvulum</i> Kütz. (Kütz.)	HANAMP	1	—	—	—	—	—	—	1	—	—	3	4	—	3	4	3	4	1	4	32	18.6				
<i>Gyrosigma acuminatum</i> (Kütz.) Rabenh.	MRDCIR	—	—	—	—	—	—	—	2	—	—	—	—	—	—	—	—	—	—	—	25	14.5				
<i>Meridion circulare</i> (Grev.) Ag.	NAVHEI	—	—	—	—	—	—	—	1	—	—	—	9	9	—	2	10	—	—	—	32	18.6				
<i>Navicula heimansi</i> Van Dam et Kooyman	NAVPU	—	—	—	—	—	—	—	—	1	—	—	5	11	9	1	1	3	—	—	31	18.0				
<i>Navicula pupula</i> Kütz.	NAVRAD	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	23	13.4				
<i>Navicula radiosus</i> Kütz.		—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	6	3.5				
<i>Neidium affine</i> (Ehrenb.) Pfitz.	NEIAFF	—	—	—	1	6	—	1	1	1	1	—	5	6	—	1	4	—	1	28	16.3					
<i>Neidium bisulcatum</i> (Lagerst.) Cleve	NEIBIS	—	—	—	—	—	—	—	—	1	—	—	4	—	7	2	—	1	5	—	1	22	12.8			
<i>Neidium hercynicum</i> A. Mayer		—	—	—	—	—	—	—	—	3	—	6	—	2	7	8	7	4	—	4	1	42	24.4			
<i>Neidium iridis</i> (Ehrenb.) Cleve		—	—	—	—	—	—	—	—	—	—	—	—	3	—	1	—	—	4	2.3						
<i>Nitzschia recta</i> Hantzsch	NITPAL	1	—	—	1	2	6	—	—	—	4	1	—	11	9	—	3	—	—	—	1	6	3.5			
<i>Nitzschia palea</i> (Kütz.) W. Smith		—	—	—	—	—	—	—	—	3	1	2	—	4	3	3	4	6	—	1	27	15.7				
<i>Pinnularia divergens</i> W. Smith	PININT	—	—	—	—	—	—	—	—	1	5	—	2	1	—	—	—	—	—	—	9	5.2				
<i>Pinnularia interrupta</i> W. Smith	PINMIC	1	—	—	7	7	6	—	—	3	9	7	6	11	10	2	6	9	—	2	86	50.0				
<i>Pinnularia subcapitata</i> Greg.	PINSUB	5	4	5	4	4	7	6	1	7	7	3	9	7	8	11	10	4	8	8	2	120	69.8			
<i>Pinnularia viridis</i> (Nitzsch.) Ehrenb.	PINVIR	7	3	5	1	7	7	6	3	7	7	9	8	8	8	8	11	10	10	9	10	9	11	90.7		
<i>Stauroneis anceps</i> Ehrenb.	STNANC	—	—	—	—	6	6	—	—	—	—	—	2	6	4	—	3	10	—	2	39	22.7				
<i>Stauroneis phoenicenteron</i> (Nitzsch.) Ehrenb.	STNPHO	2	—	—	1	5	6	—	—	—	1	—	3	11	9	—	1	9	—	—	48	27.9				
<i>Stauroneis smithii</i> Grunov	STNSMI	—	—	—	—	—	—	—	—	—	—	—	1	7	—	—	2	—	10	5.8						
<i>Stephanodiscus cf. alpinus</i> Hust.	SURANG	—	—	—	—	—	—	—	—	—	1	2	11	5	—	1	8	—	—	28	16.3					
<i>Suriella angusta</i> Kütz.		—	—	—	—	—	—	—	—	—	—	—	4	—	—	—	—	—	—	4	2.3					
<i>Suriella biseriata</i> Bréb.	SURMIN	—	—	—	—	—	—	—	—	—	—	—	—	5	3	—	—	3	—	—	11	6.4				
<i>Suriella minuta</i> Bréb.		7	1	—	7	6	5	1	1	3	7	9	6	8	8	8	1	6	8	2	4	98	57.0			
<i>Tbellaria flocculosa</i> (Roth) Kütz.	CHAOBT	—	—	—	—	—	—	—	—	—	—	—	—	1	8	—	6	—	—	15	8.7					
<i>Xanthophyceae:</i>		—	—	—	—	—	—	—	—	—	—	—	—	1	—	—	1	—	—	1	—	1	0.6			
<i>Characiopsis obtusa</i> Ettl		—	—	—	—	—	—	—	—	—	—	—	—	1	—	2	—	—	—	—	3	1.7				
<i>Characiopsis sphagnicola</i> Pascher		—	—	—	—	—	—	—	—	—	—	—	—	1	—	2	—	—	—	—	—					
<i>Characiopsis</i> spp.		—	—	—	—	—	—	—	—	—	—	—	—	1	—	2	—	—	—	—	—	—				

Sampling site (number of samples taken)	Code	(1)	(2)	(3)	(4)	(5)	(6)	(7)	(8)	(9)	(10)	(11)	(12)	(13)	(14)	(15)	(16)	(17)	Total (172)	Total (%)
<i>Ophiocytium capitatum</i> Wolle	OPHCAP	—	—	—	—	—	—	—	—	—	1	8	—	2	—	—	—	—	11	6.4
<i>Ophiocytium cochlare</i> (Eichwald) A. Braun	OPHLAG	—	—	—	—	—	—	—	—	—	2	—	—	2	—	—	—	—	8	4.7
<i>Ophiocytium lagerheimii</i> Lemmerm.	OPHPAR	—	—	—	—	—	—	—	—	—	1	—	—	9	2	—	15	8.7		
<i>Ophiocytium parvulum</i> (Perty) A. Braun	TRISPI	—	—	—	—	—	—	—	—	—	3	1	—	4	—	—	10	5.8		
<i>Tribonema regularae</i> Pascher	TRISPI	—	—	—	—	—	—	—	—	—	1	4	—	9	—	—	14	8.1		
<i>Tribonema subtilissimum</i> Pascher	TRISPI	—	—	—	—	—	—	—	—	—	1	1	—	5	—	—	7	4.1		
<i>Tribonema</i> spp.	TRISPI	—	—	—	—	—	—	—	—	—	2	1	—	—	—	—	4	2.3		
<i>Haptophyceae</i> :		—	—	—	—	—	—	—	1	—	—	—	—	1	—	1	—	3	1.7	
<i>Pseudodendromonas vlikii</i> (Vlk) Bourr.		—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	
<i>Euglenophyceae</i> :		—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	
<i>Anisonema</i> spp.	ASTCUR	—	—	1	1	1	—	—	—	—	3	—	—	3	1	2	12	7.0		
<i>Astasia curvata</i> Klebs	ASTCUR	3	—	2	2	4	—	—	2	1	2	1	—	7	—	1	25	14.5		
<i>Astasia</i> spp.	ASTCUR	4	—	—	1	2	—	1	—	—	1	2	5	1	1	21	12.2			
<i>Euglena adhaerens</i> Matv.	EUGADH	4	—	—	—	—	3	—	4	6	2	2	7	—	5	4	2	42	24.4	
<i>Euglena mutabilis</i> Schmitz	EUGMUT	3	1	—	—	1	2	3	5	4	2	4	4	—	1	3	—	3	41	23.8
<i>Euglena limnophila</i> Lemmerm.	EUGLIM	4	6	5	4	—	—	—	—	—	—	—	—	—	—	—	—	20	11.6	
<i>Euglena spirogyra</i> Ehrenb.	EUGSPI	—	—	—	—	—	—	—	—	—	6	5	—	2	—	—	13	7.6		
<i>Euglena</i> spp.	EUGSPI	—	—	—	—	—	—	—	3	—	1	1	2	—	1	5	—	13	7.6	
<i>Heteronema acus</i> (Ehrenb.) Stein	HETACU	3	3	3	3	1	1	—	—	—	—	—	—	1	—	—	15	8.7		
<i>Menodium pellucidum</i> Perty	PETMED	—	—	—	—	1	—	—	—	—	—	—	—	4	—	—	5	2.9		
<i>Petalomonas medicocanellata</i> Stein	PETMED	—	—	—	—	1	—	—	3	—	5	—	2	8	—	19	11.0			
<i>Petalomonas stetini</i> Klebs	PETMED	—	—	—	—	—	—	—	—	—	—	—	1	—	—	2	1.2			
<i>Phacus curvicauda</i> Svirenko	PHACUR	—	—	—	—	—	—	—	—	—	2	8	—	3	—	13	7.6			
<i>Phacus pyrum</i> Ehrenb.	PHACUR	—	—	—	—	—	—	—	—	—	1	—	—	2	—	—	3	1.7		
<i>Rhabdonomas costata</i> (Korshikov) Pringsh.	RHACOS	2	5	5	5	2	2	2	—	1	2	—	1	—	1	2	—	2	32	18.6
<i>Rhabdonomas incurva</i> Fres.	RHACOS	—	—	—	—	1	—	—	—	2	—	3	—	3	—	9	5.2			
<i>Trachelomonas hispida</i> (Perty) Stein em. Deflandre	TRAHIS	—	—	—	—	—	—	—	—	—	7	1	1	—	4	—	13	7.6		
<i>Trachelomonas oblonga</i> Lemmerm.	TRAOBL	—	—	—	—	—	—	—	—	—	9	6	—	9	2	—	26	15.1		
<i>Trachelomonas volvocinopsis</i> Svirenko	TRAVOL	—	1	—	4	1	4	—	—	7	3	7	4	—	1	2	3	30	23.3	
<i>Trachelomonas</i> spp.	TRAVOL	—	—	—	—	—	—	—	—	—	—	—	1	—	—	1	—	2	1.2	
<i>Tropidoscyphus octocostatus</i> Stein	—	1	—	—	—	—	—	—	—	—	1	—	—	1	1	—	5	2.9		
<i>Prasinophyceae</i> :		—	—	—	—	—	—	—	—	—	—	—	—	2	—	—	2	1.2		
<i>Monomastix pistostigma</i> Scherffel		—	—	—	—	—	—	—	—	—	—	—	—	3	—	—	5	2.9		
cf. <i>Pedinomonas minutissima</i> Skuja		2	—	—	—	—	—	—	—	—	—	—	—	—	—	—	1	—	1	0.6
<i>Scourfieldia cordiformis</i> Takeda		—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	
<i>Chlorophyceae</i> :		—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	
<i>Ankistrodesmus fusiformis</i> Corda	ANKFUS	—	—	—	—	5	—	—	—	—	1	3	—	—	10	—	19	11.0		
<i>Binucleata tectorum</i> (Kütz.) Beger in Wichmann	BINTEC	5	—	1	6	1	1	7	2	1	2	—	1	8	3	—	9	58	33.7	
<i>Botryosphaerella sudetica</i> (Lemmerm.) Silva	CHLSP1	—	—	—	—	—	—	—	2	—	7	—	—	2	—	—	11	6.4		
<i>Bulbochaete</i> sp.	CHLSP1	—	—	—	—	—	—	—	—	—	—	—	—	1	4	5	2.9			
<i>Carteria</i> spp.	CHLSP1	—	—	—	—	—	—	2	1	—	—	—	—	—	1	—	4	2.3		
<i>Chaetophora</i> sp.	CHLSP1	—	—	—	—	—	—	—	—	—	—	5	—	—	5	2.9				
<i>Chlamydomonas rotula</i> Playfair	CHLSP1	—	—	—	4	3	1	—	—	—	3	3	5	—	3	—	22	12.8		
<i>Chlamydomonas</i> sp. 1	CHLSP1	—	—	—	1	—	1	3	3	1	—	2	2	4	1	1	—	20	11.6	
<i>Chlamydomonas</i> sp. 2	CHLSP1	1	—	—	1	—	1	—	1	—	5	2	1	—	1	—	15	8.7		
<i>Chlamydomonas</i> sp. 3	CHLSP1	—	—	—	2	—	1	—	—	1	—	1	—	2	1	—	9	5.2		
<i>Chlamydomonas</i> sp. 4	CHLSP1	—	—	—	1	—	—	—	—	—	—	—	—	4	—	—	5	2.9		
<i>Chlamydomonas</i> spp.	CHLSP1	1	2	—	1	2	2	3	—	3	5	3	1	4	4	3	3	52	30.2	
<i>Chlorella</i> spp.	CHLMSP	—	—	—	—	—	—	—	—	—	—	—	1	—	—	—	—	3	4.2	
<i>Chlorococcum</i> sp.	CHLMSP	—	—	—	2	—	—	—	—	—	—	—	—	—	—	—	—	2	1.2	
<i>Chlorogonium elongatum</i> Dang.	CHLMSP	—	—	—	—	—	—	—	—	—	—	—	1	—	—	3	—	4	2.3	
<i>Chlorogonium</i> sp.	CHLMSP	—	—	—	2	—	—	2	—	1	—	1	1	—	1	2	11	6.4		
<i>Chloromonas</i> sp. 1	CHLMSP	1	6	5	5	—	—	—	—	—	—	—	—	—	—	—	—	17	9.9	
<i>Chloromonas</i> spp.	CHLMSP	—	—	—	—	—	—	—	—	—	2	1	—	2	—	—	6	3.5		
<i>Choricystis chodatii</i> (Jaag) Fott	COCLAC	—	—	—	1	—	1	—	—	—	—	—	—	—	—	—	—	2	1.2	
<i>Coccomyxa confluens</i> (Kütz.) Fott	COESPH	—	—	—	2	—	1	—	1	3	—	—	2	—	1	1	11	6.4		
<i>Coccomyxa lacustris</i> (Chodat) Pascher	COESPH	2	—	—	3	2	—	—	7	—	—	1	7	5	—	3	30	17.4		
<i>Coenochlors sphagnicola</i> Hindák	COESPH	—	—	—	1	—	—	—	—	—	—	—	—	—	—	—	—	—		
<i>Dicranochaete reniformis</i> Hieron.	COESPH	7	—	—	—	4	—	1	—	—	—	—	—	4	3	—	19	11.0		
<i>Dictyosphaerium sphagnale</i> Hindák	COESPH	—	—	—	—	—	—	6	—	—	—	—	2	—	—	8	4.7			
<i>Gloeocystis polydermatica</i> (Kütz.) Hindák	COESPH	—	—	—	—	—	—	—	—	—	—	—	—	—	1	—	1	0.6		
<i>Gonium sociale</i> (Dujard.) Warming	COESPH	—	—	—	—	—	—	—	—	—	—	—	—	1	—	—	3	1.7		
<i>Lobomonas rostrata</i> Hazen	COESPH	—	—	—	—	—	—	—	—	—	—	—	—	3	—	—	3	1.7		
<i>Microspora quadrata</i> Hazen	COESPH	1	—	—	—	2	1	—	—	—	—	1	—	—	5	—	—	5	2.9	
<i>Microspora stagnorum</i> (Kütz.) Lagerh.	COESPH	1	—	—	—	2	3	1	1	—	—	3	—	—	1	—	13	7.6		
<i>Microspora</i> spp.	COESPH	2	—	—	1	1	—	1	1	1	2	1	—	—	3	—	3	17	9.9	
<i>Oedogonium</i> sp.	OEDOSP	6	—	—	7	2	2	6	—	—	3	—	5	3	—	1	5	10	59	34.3
<i>Oocystis solitaria</i> Wittr.	OOC SOL	7	—	—	2	1	—	8	—	8	5	4	—	2	7	2	11	10	68	39.5
<i>Oocystis</i> spp.	OOC SOL	—	—	1	—	—	2	—	—	—	—	1	1	—	3	2	10	5.8		
<i>Palmodictyon varium</i> (Näg.) Lemmerm.	PALVAR	—	—	—	3	6	2	—	—	—	8	—	—	—	—	—	19	11.0		

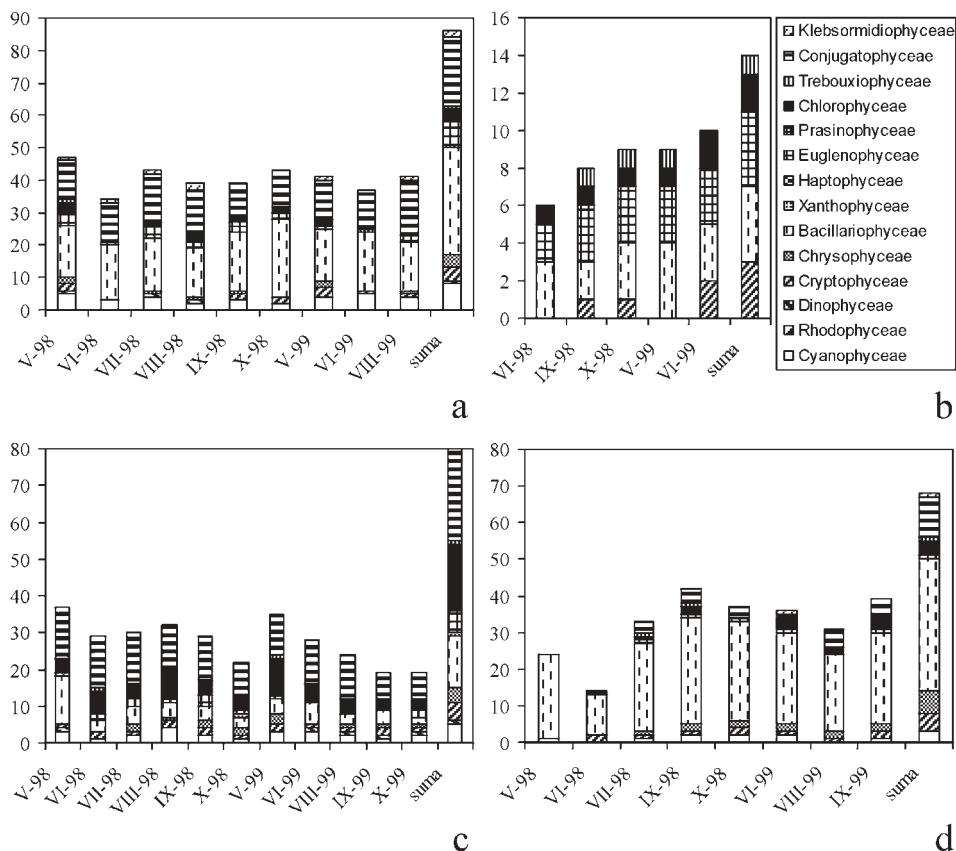


Fig. 1.—Composition of algal flora in different types of waterbodies in 1998–1999: a – bog lake (UR-20), b – intensively shaded pool (PR-3), c – shallow peaty pool (PR-12), d – loamy-bed pool (PR-14). PR – Pančavské rašeliniště, UR – Úpské rašeliniště.

Besides ubiquitous species such as *Achnanthes minutissima*, *Pinnularia subcapitata* and *P. viridis*, many typical acidophilic and oligotrophic species were found [*Chroococcus turgidus*, *Synura sphagnicola*, *Eunotia rhomboidea*, *Botryosphaerella sudetica*, described by Lemmerman (1896) from Úpské rašeliniště, *Oocystis solitaria*, *Closterium nilssonii*, *Euastrum humerosum*, *E. insigne*, *Netrium digitus*, *Penium polymorphum* etc.], psychrophilic species (e.g. *Eunotia glacialis*, *Pinnularia lata*, *Pedinomonas minutissima*, *Scourfieldia cordiformis*, *Binuclearia tectorum*) or species with arctic-alpine distribution (*Anomoeoneis brachysira*, *Cymbella hebridica*, *Frustulia rhombooides*, *Pinnularia lata*, *Penium cylindrus*). The occurrence of these species corresponds well to the type of the locality (subarctic-subalpine peat bog). On the other hand some eutrophic species (*Cyclotella radiososa*, *Heteronema acus*, *Surirella angusta*, *Scenedesmus acutus*, *S. ecornis*) were recorded, too.

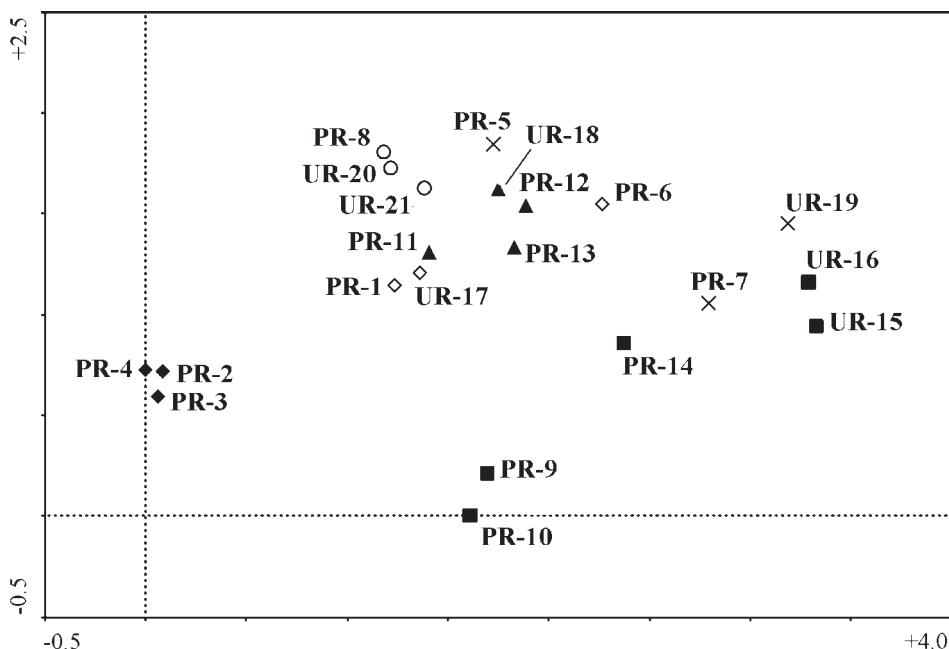


Fig. 2. – DCA ordination diagram showing the position of localities; ○ – bog lakes (Blänk or Kolk in German), ♦ – intensively shaded pools, ◇ – partly shaded pools, ▲ – shallow peaty pools, ■ – pools with loamy bed, × – others (not classified). PR – Pančavské rašeliniště, UR – Úpské rašeliniště.

The location of the pools in particular bogs did not influence the algal flora; differences in species composition were caused by the character of the pools. On DCA plot (Fig. 2), pools of a similar type form clusters. Four categories could be distinguished in accordance with the morphology of pools and composition of their algal flora (Figs 1, 2):

1. Bog lakes (“Blänk” or “Kolk” in German, Fig. 1a). Planktonic species prevail, mainly the members of *Zygnematophyceae* and *Chlorophyceae*; the diatoms (*Bacillariophyceae*) were less represented.
2. Shallow peaty pools (Fig. 1c) had the most diversified algal flora with the dominance of desmids (*Desmidiales*, *Zygnematophyceae*) and diatoms.
3. Intensively shaded pools (Fig. 1b) comprised very poor communities
4. Loamy-bed pools (deep waterbodies, pools in streams, Fig. 1d). In spite of great differences in the species composition due to dissimilar pH, a high proportion of the diatoms is characteristic for these biotopes.

Algal flora of the partly shaded pools shared features of shallow peaty pools and intensively shaded pools; some other water bodies did not belong to any of these categories.

CCA demonstrated pH, shading and type of bed as the most important factors influencing the diversity of algae; pH affected especially the species composition (Figs 3a, b). This effect was most evident in the presence of the *Xanthophyceae*, which were found only in the pools with higher pH values, and in the occurrence of the dominant *Bacillariophyceae*: *Frustulia*

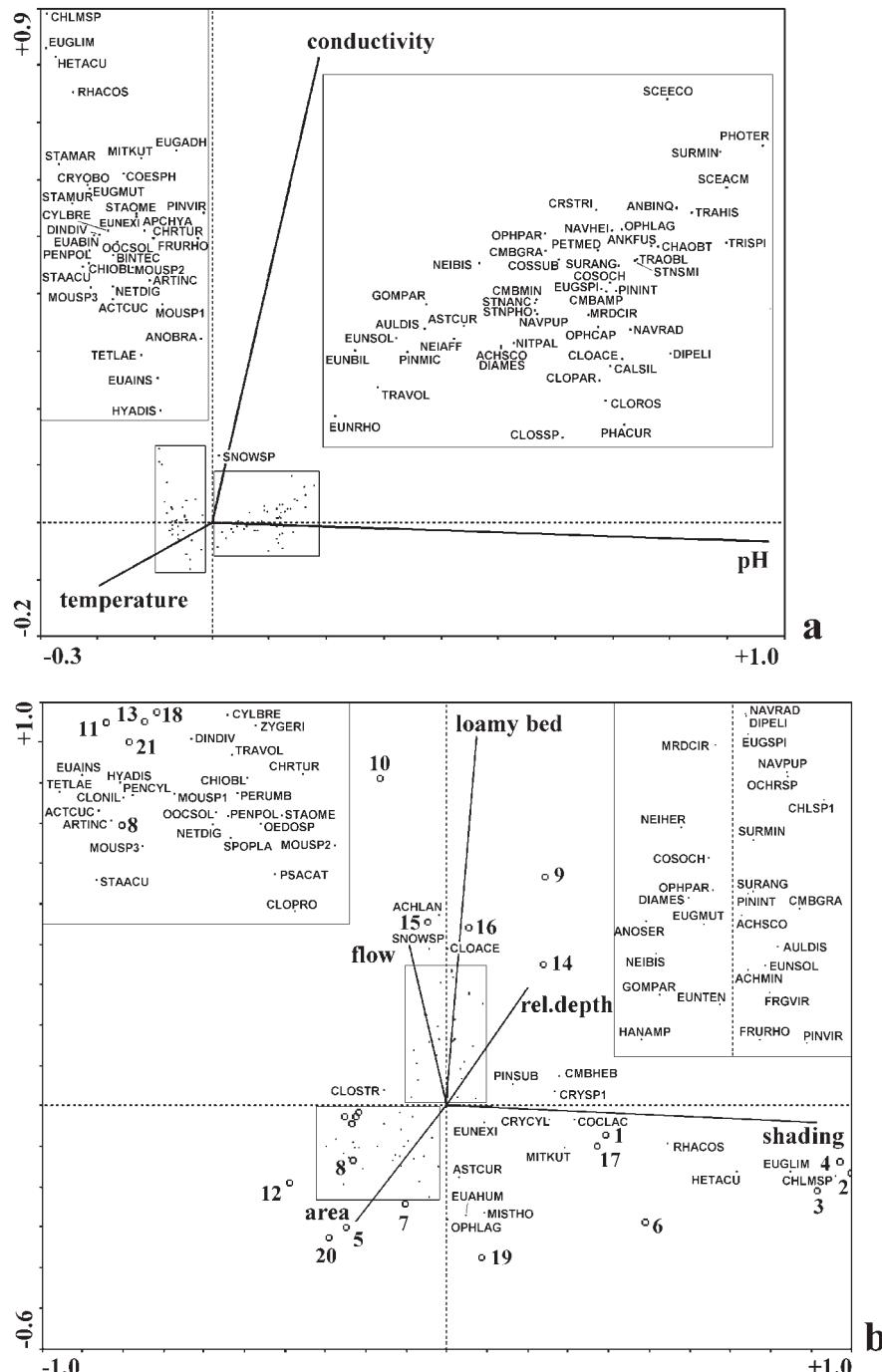


Fig. 3. –CCA ordination diagrams showing the position of species with respect to: a – physical and chemical water parameters, b – morphological characteristics of the pools. See Table 2 for species codes. Only well fitting species are displayed.

rhomboides was the most abundant diatom in the acid pools with average pH < 5. Where pH varied around 5, *F. rhomboides* shared the dominance with *Fragilaria virescens*, and in more alkaline pools, *F. virescens* and representatives of the genus *Navicula* dominated. The number of species increased with rising pH-values as well. On the other hand, shading decreased the number of species. The presence of few unique species (*Euglena limnophila*, *Heteronema acus*, *Rhabdomonas costata*, *Chloromonas* sp. 1), together with some common taxa (*Pinnularia subcapitata*, *Cryptomonas* sp. div.), was typical of the intensively shaded pools. The type of bed determined the dominant group of algae.

The group of species in the bottom left corner of Fig. 3b seems to correlate remarkably with the variable “area”, but in fact, it reflects a negative position to the variables “shading” and “loamy bed”. Majority of these species are similar to the left, “acidophilic” cluster in Fig. 3a and it is exactly these species that could be considered as typical for the peat bogs.

Discussion

It is difficult to compare the present occurrence of algae with former records from the same area because of the investigators’ unequal interest in various groups of algae and taxonomic changes. For example Lemmermann (1896) did not determine the diatoms, Beck-Managetta (1927) dealt only with *Cyanophyceae*, *Chlorophyceae* and *Zygnematophyceae*. Most of the dominant algae were finally identified, and their occurrence was confirmed. However, lot of species (some of them rather distinctive) have not been found. This need not to be necessarily caused by the local extinction of these species but could reflect the fact that they become abundant only in certain years. Another reason could be the spatial heterogeneity of algal communities. Some species were found only in one or a few pools. Considering the fact that the number of the observed pools represents only a small part of the pools in the peat bogs, further research could bring new results.

One hundred and ninety taxa of algae were found in the samples from the Úpské rašeliniště peat bog. Matula (1995) encountered 114 species of algae in the same locality during his investigation. Although the overall portion of the three dominant groups (*Bacillariophyceae*, *Chlorophyceae*, *Zygnematophyceae*) was in both cases about 70%, their individual ratios differ distinctly (Table 3). Unfortunately no details are available as I was interested more in the differences between the bogs than between the microbiotopes within particular bogs.

Table 3. – Comparison of the composition of algal flora from the Úpské rašeliniště peat bog as recorded by Matula (1995) and in the present study.

Author	Matula 1986–90	Nováková 1998–99
Period of data collation		
Total number of taxa	114	190
<i>Bacillariophyceae</i> (%)	13	31
<i>Chlorophyceae</i> (%)	26	18
<i>Zygnematophyceae</i> (%)	31	19
others %	30	32

The ecology of algae in the Czech mountain bogs was studied in the Jizerské hory Mts and in the Šumava Mts. Gessner (1933) and Perman (1961) dealt with plankton production according to nutrient content and other physico-chemical factors in peat bog lakes in the Jizerské hory Mts. Perman (1961) found the chrysophytes, the cryptomonads and also the diatoms dominating in plankton. The high diatom abundance is in contradiction with the results of Gessner (1933) who referred to diatoms as being in minority. Although the diatoms dominated in most samples from the Krkonoše Mts bogs, they were found only in small quantities in larger lakes (PR-8, UR-20) where plankton formed the main part of algal flora.

Lederer (1997, 1998) investigated several peat bogs in the Šumava Mts and concentrated on the ecology of algae found in various aquatic and subaerophytic microbiotopes. He concluded that algal flora of particular microbiotopes varied more than between various localities (peat bogs). This conclusion is in agreement with my results. Lederer found 193 taxa of algae in the Šumava Mts bogs, but among these only 26 species (13%) were diatoms. However, he determined an unusually high number of *Cyanophyta* (30 species); some of these taxa were rare and one species was newly described (Lederer 1995a, 1995b).

It should be questioned to what extent these differences in proportions of algal classes are caused by real dissimilarities in algal communities in various bogs and to what extent by the authors' interest in different classes of algae. This fact is evident in most of the papers dealing with the ecology of algae in other European bogs, which concentrate only, or mainly, on desmids (e.g. Péterfi 1974, Wurm 1991, Wurm & Krisai 1993, Tomaszewisz 1994).

Acknowledgements

I am grateful to Tomáš Kalina and Yvonne Němcová for their valuable remarks to the manuscript. To Eric Engstrom I am indebted for linguistic corrections. The work was supported by the grant no. 206/98/1193 from the Grant Agency of Czech Republic and research grant no. J13/98113100004 from the Czech Ministry of Education.

Souhrn

Během dvouletého výzkumu tůní Úpského a Pančavského rašelinistě v Krkonoších bylo nalezeno 228 taxonů silnic a řas. Nejhojnější skupinou byly rozsivky, následované krásivkami. Poloha sledovaných tůní v různých rašelinistických neměla vliv na druhové složení řas, rozdíly v algoflóře byly dány rozdílným charakterem stanovišť. Studium vztahů mezi algoflórou a proměnnými prostředí ukázalo, že faktory nejvýznamněji ovlivňující řasová společenstva byly pH, zastínění a charakter dna tůně.

References

- Beck-Managetta G. (1927): Algenfunde im Riesengebirge. – Věstník Král. Čes. Spol. Nauk, ser. mat.-nat., Praha, (1926): 1–18.
- Beck-Managetta G. (1929): Algenfunde im Riesengebirge. – Lotos, Praha, 77: 92–100.
- Gessner F. (1933): Nährstoffgehalt und Planktonproduktion in Hochmoorblänken. – Arch. Hydrobiol., Stuttgart, 25: 394–404.
- Jeník J. & Soukupová L. (1992): Microtopography of subalpine mires in the Krkonoše Mountains, the Sudets. – Preslia, Praha, 64: 313–326.
- Kalina T. (1969a): *Gloeochrysis montana* n. sp. und *Poterioochromonas stipitata* Scherffel (*Chrysophyceae*) aus Krkonoše (Riesengebirge). – Österr. Bot. Z., Wien, 117: 139–145.
- Kalina T. (1969b): Submicroscopic structure of silica scales in some *Mallomonas* and *Mallomonopsis* species. – Preslia, Praha, 41: 227–228.
- Kalina T. (1970): *Corcontochrysis noctivaga* gen. et sp. n. (*Chrysophyceae*). – Preslia, Praha, 42: 297–302.

- Kramer K. & Lange-Bertalot H. (1986): *Bacillariophyceae*, 1. Teil *Naviculaceae*. – In: Ettl H., Gärtner G., Heynig H. & Mollenhauer D. (eds.), Süßwasserflora von Mitteleuropa, G. Fischer Verlag, Jena.
- Lederer F. (1995a): A new species of *Cyanodictyon* (*Cyanoprokaryota, Chroococcales*) from peat-bogs in the Šumava Mts, Czech Republic. – Preslia, Praha, 67: 117–121.
- Lederer F. (1995b): Several little known *Cyanobacteria/Cyanoprokaryota* from peat-bogs in the Šumava Mts, Czech Republic. – Arch. Hydrobiol. Suppl. 111/Algol. Stud., Stuttgart, 79: 57–65.
- Lederer F. (1997): Řasová flóra šumavských rašeliníšť. – Erica, Plzeň, 6: 3–14.
- Lederer F. (1998): Srovnání mikroflóry rašeliníšť Šumavy a Třeboňské páne. – Ms., 98 pp. [Kandid. dis. pr.; depon. in Knih. Bot. Úst. AV ČR, Třeboň].
- Lemmermann E. (1896): Zur Algenflora des Riesengebirges. – Forschb. Biol. Stat., Plön, 4: 88–133.
- Matula J. (1980a): Algae new or rare to the Polish flora, found in peat bogs of the Sudeten Mts. – Fragm. Flor. Geobot., Kraków, 26: 121–136.
- Matula J. (1980b): *Ducellieria chodatii* (Ducel.) Teil. and *D. corcontica* Mat. nova sp. (*Xanthophyceae*) found in a peat bog in the Karkonosze Mts. – Fragm. Flor. Geobot., Kraków, 26: 349–353.
- Matula J. (1995): Warunki troficzne glonów torfowiskowych na obszarze Dolnego Śląska. – Wydaw. Akad. Rolnicz., Wrocław.
- Mejstřík V. & Straka K. (1964): Pančické rašeliníště v Krkonoších. – Opera Corcontica, Praha, 1: 35–53.
- Müller O. (1898): *Bacillariales* aus den Hochseen des Riesengebirges. – Forschb. Biol. Stat., Plön, 6: 48–82.
- Němcová Y., Kalina T., Neustupa J. & Nováková S. (2001): Silica-scaled chrysophytes of the Krkonoše Mts (Czech Republic) – Arch. Hydrobiol. Suppl. 137/Algol. Stud., Stuttgart, 101: 97–108.
- Perman J. (1961): Řasová flóra některých dystrofních vod v Jizerských horách. – Severočeské Muzeum, Liberec.
- Péterfi L. S. (1974): Structure and patterns of desmid communities occurring in some Romanian peat bogs. – Nova Hedwigia, Stuttgart, 25: 651–664.
- Pochmann A. (1940): Mikrofloristisches Streifzug im Riesengebirge. – Mikrokosmos 7: 94–109.
- Rudolph K., Firbas F. & Sigmund H. (1928): Das Koppenplanmoor im Riesengebirge. – Lotos, Praha, 76: 173–222.
- Schröder B. (1898): Neue Beiträge zur Kenntnis der Algen des Riesengebirges. – Forschb. Biol. Stat., Plön, 6: 9–47.
- ter Braak C. J. F. & Šmilauer P. (1998): CANOCO references manual and user's guide to Canoco for Windows. Software for Canonical Community Ordination (version 4). – PRO-DLO, Wageningen.
- Tomaszewicz G. (1994): Abundance and composition of the desmid flora in a series of peat pits, in relation to pH and some other habitat parameters. – Biológia, Bratislava, 49: 519–524.
- van den Hoek C., Maan D. G. & Jahns H. M. (1995): Algae – an introduction to phycology. – Cambridge University Press, Cambridge.
- Wurm E. (1991): Rezente und subfossile Algenvegetation. – In: Krisai R., Burgstaller B., Ehmer-Künkele U., Schiffer R. & Wurm E. (eds.), Die Moore des Ost-Lungau. Heutige Vegetation, Entstehung, Waldgeschichte ihrer Umgebung, Sauteria, Salzburg, 5: 53–108.
- Wurm E. & Krisai R. (1993): Schrenkenbüchelmoos und Konradenmoos, zwei Fichtenmoore in den östlichen Zentralalpen. – Mitt. Abt. Bot. Landesmus. Joanneum Graz 21/22: 55–94.
- Zacharias O. (1896): Ergebnisse einer biologischen Excursion an die Hochseen des Riesengebirges. – Forschb. Biol. Stat., Plön, 4: 65–87.
- Zacharias O. (1898): Summarischer Bericht über die Ergebnisse meiner Riesengebirgsexcursion von 1896. – Forschb. Biol. Stat., Plön, 6: 1–8.

Received 1 November 2000
Revision received 21 July 2001
Accepted 27 September 2001